Nitrogen status regulates morphological adaptation of marsh plants to elevated CO₂

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Coastal wetlands provide valuable ecosystem services that are increasingly threatened by anthropogenic activities¹. Atmospheric carbon dioxide (CO₂) concentration has increased from 280 to 404 ppm since the industrial revolution, and is projected to exceed 900 ppm by 2100². In terrestrial ecosystems, elevated CO₂ typically stimulates C₃ plant photosynthesis and primary productivity leading to an increase in plant size³. However, compared to woody plants or crops⁴, the morphological responses of clonal non-woody plants to elevated CO₂ have been less well-studied. We show that 30 years of experimental CO2 enrichment in a brackish marsh increased primary productivity and stem density, but decreased the stem diameter and height of the dominant clonal species Schoenoplectus americanus. Smaller, denser stems were associated with the expansion of roots and rhizomes to alleviate nitrogen (N) limitation as evidenced by high N immobilization in live tissue and litter, high tissue C:N, and low available porewater N. Changes in morphology and tissue chemistry induced by elevated CO₂ were reversed by N addition. We demonstrate that morphological responses to CO₂ and N supply in a clonal plant species influences the capacity of marshes to gain elevation at rates that keep pace with rising sea levels.

Terrestrial plants are experiencing the highest atmospheric CO₂ concentration in the past 800,000 years⁵. The stimulation of carbon (C) fixation by elevated CO₂ -- the CO₂ fertilization effect -- is well documented⁶. Increases in leaf-level C uptake rates are often accompanied by changes in plant morphology or morphometric sizes, such as increased height, stem diameter, leaf area index, leaf number, and root volume^{7, 8, 9, 10}. These morphometric changes are widely observed to influence competitive dynamics^{11, 12}, with implications for ecosystem structure and function.

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Resource availability regulates the effect of elevated CO₂ on total biomass production but can also influence changes in plant size and patterns of biomass allocation. Elevated CO₂induced changes in plant morphology are poorly understood compared to changes in total biomass, and morphological changes are highly variable along environmental gradients and between plant functional groups, reflecting species-specific strategies for resource acquisition⁷, 9. For instance, plant species can overcome N limitation caused by CO₂ fertilization in low-N environments if they possess traits to mine and recycle N through shared underground networks of ectomycorrhizae¹³, illustrating the importance of understanding how resource acquisition influences plant responses to elevated CO₂. Similarly, clonal plants employ shared underground networks of roots and rhizomes to exploit heterogeneous soil resources, affording a competitive advantage in resource-poor environments¹⁴. Clonal plants are ubiquitous, occurring in 10 out of 11 classes of vascular plants, and are an important component of many ecosystems¹⁵; yet, their morphological responses to elevated CO₂ are understudied compared to other functional groups such as non-clonal trees, shrubs, herbs, and crops. Clonal architecture has important implications for biomass allocation and resource acquisition strategy that can propagate into ecosystem level responses to global change. For example, growth allocation to rhizomes and ramets may influence stem density and decrease interannual variation in plant growth by storing resources when conditions are favorable and remobilizing them when conditions are adverse¹⁴.

Our objective was to examine the morphological responses of a clonal marsh plant species to elevated CO₂ and N addition using long-term data from two field experiments in a tidal marsh on the Chesapeake Bay. The native plant community is dominated by the perennial C₃ sedge *Schoenoplectus americanus* and two co-dominant C₄ grasses, *Spartina patens* and *Distichlis spicata*. *Experiment 1* began in 1987 and consists of open-top chambers ventilated with either ambient air or CO₂ concentrations elevated to 700-800 ppm. *Experiment 2* began in 2006 to investigate interactions between elevated CO₂ and N addition with 4 treatments: ambient CO₂, ambient CO₂ + N, elevated CO₂, and elevated CO₂ + N. In both experiments, we made annual measurements of stem morphology (height, width, and density), stem biomass, belowground productivity, tissue chemistry, porewater ammonium (NH₄⁺, started in 2002 in *Experiment 1*), and soil surface elevation change. Morphometric data were collected only for *S. americanus*; the morphometric responses of C₄ grasses were not examined because the growth form of these species does not lend itself to such measurements¹⁶.

Elevated CO_2 increased sedge total biomass by an average of 20% over the control in Experiment 1. The CO_2 -driven increase in belowground biomass productivity (34% \pm 7) was larger than the increase in aboveground biomass (17% \pm 4), resulting in a 16% \pm 6 increase in the below:above biomass ratio (Fig. 1a). Simultaneously, the density of stems increased 51% \pm 6 (Fig. 2a, Appendix 1, Table S1) and the biomass of individual sedge stems decreased $16\% \pm 1$ (Fig. 2b), corresponding to a 5% decline in stem height and a 10% decline in diameter (Fig. 2c, d). Moreover, stem density and rhizome biomass were positively correlated ($R^2=0.30$, P < 0.0001, Fig. S1), suggesting that the CO_2 -induced increase in belowground allocation was expressed through the clonal architecture of the sedge.

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Plants shift biomass allocation between roots and shoots to optimize resource capture and use, with allocation to photosynthetic aboveground biomass for CO₂ uptake, and to belowground biomass for nutrient uptake¹⁷. Salt and brackish systems tend to be N limited because the phosphorus that would be bound to iron in freshwater systems is liberated by the sulfates in seawater¹⁸. N limitation is known to constrain the CO₂ fertilization effect^{13, 19, 20}, and in our experiments elevated CO₂ induced the classic symptoms of progressive N limitation in this coastal wetland ecosystem¹⁹. Elevated CO₂ plots in Experiment 1 had more N sequestered in plant biomass (i.e., 5% increase in shoot N and 12% increase in root N) and litter (21% increment, Fig. 1a), and less plant-available inorganic soil N (i.e., 47% lower porewater [NH₄+]; [NO₃⁻] was below detection, Fig. 1a, Fig. S2a), implying soil N depletion in the rooting zone. Elevated CO₂ increased the C:N ratio in sedge shoot, root, and rhizome biomass by 18%, 10%, and 23%, respectively (Fig. 1a), consistent with a CO₂-induced increase in N use efficiency²¹. Preferential allocation of photosynthate to roots and the rhizome network helps alleviate N depletion by expanding the surface area for N acquisition²². In this clonal species, rhizome extension leads to increased tiller recruitment and ultimately higher stem density²³.

Experiment 2 provides an experimental test of inferences about CO_2 -induced N limitation from Experiment 1. In Experiment 2, elevated CO_2 alone produced the same response in sedge density and size observed Experiment 1, but the effect was absent when elevated CO_2 was crossed with N addition, in which case individual *S. americanus* stem size increased (Fig. 1b). As in Experiment 1, the CO_2 -only treatment decreased inorganic N by 48% \pm 4 (Fig. 1b, Fig. S2b), while adding N to elevated CO_2 plots increased porewater inorganic N by 5% \pm 9 higher than that in ambient plots. Moreover, the N-only treatment resulted in a 15% decrease in belowground productivity, indicating reduced plants biomass allocation to root systems for N uptake under N enrichment.

In contrast to the common result that plant size increases under elevated CO₂, we observed that elevated CO₂ caused a clonal plant to produce smaller individual stems at higher density. While "shrinking stems" in response to elevated CO₂ have been observed in other C₃ grasses⁸, we clearly demonstrate that this is an indirect response to N deficiency in a clonal plant species through manipulative experiments. We propose a conceptual framework for the responses of clonal plant growth to CO₂ enrichment (Fig. 3). Elevated CO₂-stimulation of plant productivity (clonal and non-clonal) leads to soil N deficiency because the increase in N demand is not satisfied by a combination of increased N uptake and shifts in plant C:N ratio. In most plant functional groups, this shift leads to an increase in growth allocation to belowground structures and increased N uptake without necessarily changing plant density. However, in clonal species, increasing belowground growth generates a more extensive rhizome system, more tiller-recruitment of stems, and increased stem density. Because the increase in stem density is not entirely compensated by higher ecosystem NPP, individual plant stems are

smaller. Because shrinking stems coincide with N depletion and can be reversed by N enrichment (Fig. 1b), we conclude that the response of clonal plant morphology to elevated CO₂ is regulated by soil N limitation (Fig. 3, Fig. S3) and suggest that shrinking stem sizes can be used as evidence of N limitation in clonal plant communities. Such responses to CO₂-induced N limitation are observed in a variety of terrestrial ecosystems where clonal species occur, but the consequences of allocation shifts for plant morphology, stand structure, and ecosystem function have been largely overlooked.

The phenotypic plasticity of marsh plants in response to elevated CO₂ has an important impact on the survival of coastal wetlands and their ecosystem services. The effectiveness of wetland vegetation in dissipating storm energy is tied to the density and morphology of stems, and their effect on frontal area (i.e., the total cross-sectional area of stems perpendicular to flow velocity)^{24, 25}. We calculated changes in frontal area in response to elevated CO₂ in *Experiment 1* and *2* and found significant increases in frontal area due to increased stem density (Table 1). The analysis indicates that elevated CO₂ may enhance the value of marshes as natural infrastructure for coastal protection through a mechanism tied to clonal traits²⁴. These morphological changes in response to elevated CO₂ also have important implications for the habitats of wetland birds, nekton and benthic invertebrates^{26, 27, 28}, and they influence key ecosystem processes such as soil formation, C storage, and nutrient retention²⁹.

Perhaps most importantly, changes in root and shoot morphology directly influence the capacity of coastal wetlands to build elevation in response to rising sea level^{30, 31}. At our high marsh site in the Chesapeake Bay, 69% of the species are clonal, and clonal plants occupy 68%

of marsh area. Tradeoffs between shoot and root productivity under elevated CO2 potentially alter the balance between the contributions of surface mineral sedimentation and subsurface root expansion to elevation gain¹. Long-term field measurements indicate that elevated CO₂enhanced belowground production increased elevation gain by 1.5 mm yr⁻¹ via subsurface expansion³². As sea level rises and plants are flooded more frequently, aboveground biomass and stem morphology also influence elevation gain by enhancing the settling of suspended mineral sediments on the marsh surface^{33, 34, 35}. We explored the effects of changes in stem structure on potential mineral deposition using a previously published model^{35, 36} to simulate increased flooding 25 years in the future (~8 cm of increased flooding, Appendix 2). Modeled surface sedimentation was enhanced an additional 0.7-1.5 mm yr⁻¹ by elevated CO₂ and the combination of elevated CO₂ and N (Table 1), suggesting an aboveground mechanism for increasing the resilience of C₃ marsh ecosystems to sea level rise. Feedbacks between aboveground biomass and mineral sedimentation become stronger with increasing flooding duration, and suspended sediment supply^{1, 35}, suggesting that the positive effects of elevated CO₂ on elevation gain may amplify under these conditions. Such responses will vary with factors such as suspended sediment supply, watershed N loading, plant traits, and other site-specific characteristics. For example, plants responded to the combination of elevated CO₂ and N in Experiment 2 with an increase in stem size, density, aboveground biomass, and belowground productivity compared to the ambient treatment, which translated into both higher modeled surface accretion and subsurface expansion (Table 1). However, N can also cause root biomass to decline³⁷, in which case elevation gains from increased surface accretion may be offset by lower subsurface expansion. Process-based models informed by observations and experiments

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are required to integrate the complex feedbacks that ultimately govern elevation gain. Plant traits such as clonal architecture that govern density, height, root allocation, and other morphological responses to global change have important ecosystem consequences that are presently missing from forecast models.

We showed that elevated CO₂ and N addition elicit significant changes in the structure and function of coastal marshes that arise from tractable plant traits such as clonal architecture. The 30-year data set presented here raises additional questions that are outside the scope of the present study. Firstly, Rasse et al. summarized the data from *Experiment 1* after 17 years and found that *S. americanus* density in the C₃ community increased by 128% compared to 51% in the present study³⁸ (Fig. 2a). Our results demonstrate that the direction of the density responses remained the same after an additional 13 years CO₂ enrichment, but the amount of stimulation has declined due to resource limitation, interactions with other variables such as sea level rise, or other factors. Secondly, the stem biomass and density responses in the C₃ community changed over time (Table S1), perhaps with changes in salinity, NH₄+ discharges from the Rhode River watershed, or other factors that require additional exploration (Table S2). Thus, future studies must consider additional important factors such as warming temperatures, changing precipitation and hydrologic extremes, changing salinity and inundation regimes, and invasive species.

- 171 Supplementary Information
- 172 Appendix 1 and Appendix 2
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- 277 Author Contributions The analysis was conceived by M.L. and J.P.M. The ongoing operation of the
- 278 experiments were conducted by J.P.M. and J.A.L. The data were compiled and analyzed by M.L.
- 279 Accretion modelling was performed by E.H. All authors contributed to writing the paper.
- 280 Author Information Reprints and permissions information is available at www.nature.com/reprints. The
- authors declare no competing financial interests. Correspondence and requests for materials should be
- addressed to M.L. (lum@si.edu) and J.P.M. (megonigalp@si.edu).

The open top chamber (OTC) CO₂ enrichment experiment, initiated in 1986, is located at Kirkpatrick marsh (38°53′N, 76°33′W), a 23-ha brackish salt marsh on the Rhode River estuary in Chesapeake Bay, on the eastern coast of the United States. This common Chesapeake Bay high marsh ecosystem is dominated by a perennial C₃ sedge *Schoenoplectus americanus*, and two co-occurring C₄ grasses *Spartina patens* and *Distichlis spicata*. Mean annual temperature is 14.1 °C and mean annual precipitation is 948 mm. Mean tidal range is approximately 40 cm, and the marsh platform is about 18 cm (in the C₃ plots) and 40 cm (in the C₄ plots) above daily mean low water level. Water level is higher than mean marsh elevation about 25% of the time. The soils contain approximately 80% organic matter (i.e. peat) to a depth of 5 m. The morphometric data used in our analysis was available only for C₃ sedges; morphometric data are not collected on C₄ grasses in these long-term experiments because of their small stem diameter and high density (Appendix 1).

Experiment 1: Three plant communities were distinguished in the marsh, a C_3 community dominated by *S. americanus*, a C_4 community consisting of *S. patens* and *D. spicata*, and a mixed C_3/C_4 community. OTC was used to increase the CO_2 concentration around the plants in each of the three communities. Ten circular chambers of 80 cm diameter and 100 cm height were placed in each community. In five of the chambers, atmospheric $[CO_2]$ was elevated to $340 \ \mu I \ CO_2 \ I^{-1}$ above the ambient CO_2 concentration (about $340 \ \mu I \ CO_2 \ I^{-1}$ at the beginning of the study). The CO_2 concentration in the other five chambers was ventilated with ambient air as the ambient treatment. To determine a possible OTC effect on plant growth, five outside chamber

control sites in each community were compared with the ambient CO_2 chambers. CO_2 exposure began each year when the plants emerged in the spring and continued 24h a day through autumn following total senescence. A survey of all plots was conducted in 1986 before initiation of the treatments and showed no significant differences in plant biomass assigned to the three treatments in each community.

Experiment 2: Twenty octagonal OTCs of 200 cm height and 150 cm diameter were established adjacent to the site of Experiment 1 in 2006. OTCs were randomly assigned to one of four treatment groups (n=5): ambient CO₂, ambient CO₂ + N, elevated CO₂, and elevated CO₂ + N. CO₂ treatment was consistent with Experiment 1. N was added by spraying NH₄Cl monthly from May to September each year (total 25 g N m⁻² year⁻¹).

Total height, green height and half height diameter of each C₃ sedge stem were measured and stem density was counted in the plots each year. The dry mass of individual stems was determined using an allometric equation based on destructively harvested samples¹⁶, and the resulting mean biomass was multiplied by the stem density to estimate aboveground biomass. Belowground plant productivity was estimated each year by three root ingrowth cores in each plot. Nine porewater wells were placed in each chamber with three depths: 15, 30 and 75 cm. Porewater was sampled and NH₄+ concentration was analyzed approximately monthly during the growing season. In these anaerobic marsh soils, porewater NO₃- is typically below detection limits and does not contribute substantially to soil available N.

In this study, we calculated the response ratio to reflect the response of plant and porewater N to elevated CO₂. Response ratio is defined as the ratio of the mean value of a given variable in

the treatment group (elevated CO₂ or elevated CO₂ plus N addition) to that in the control group. The ratio for each year was calculated from the means of replicate plots (generally n=5) in each treatment. The annual ratios were then averaged to reflect the treatment effect across the 30-year (*Experiment 1*) or 11 year (*Experiment 2*) record. Repeated measures using a mixed effects model was used to test for significant differences in stem biomass, height, diameter, and densities between the elevated CO₂ and ambient CO₂ chambers in SAS program (version 9.0). A discrete autoregressive correlation model was conducted to test time effects. The environmental and climate impact was examined with correlations between C₃ sedge aboveground biomass and annualized environmental factors using Pearson's r statistic. Figures and linear regression results were conducted using SigmaPlot (version 10.0, SPSS Inc., Chicago, IL, USA).

We modeled changes in potential mineral accretion using a previously published model of marsh vertical accretion that takes into account the height, diameter, and density of marsh vegetation^{35, 36}. We used treatment-specific relationships between aboveground biomass and stem structure (density, diameter, and height) from *Experiments 1 and 2*. Because mineral accretion rates at this high elevation marsh are presently limited by very infrequent surface flooding, we modeled the effect of biomass and stem structure on mineral accretion rates for a future marsh that is lower elevation and more frequently flooded due to accelerated sea level rise. Specifically, model experiments used a lower elevation of Kirkpatrick Marsh (0.1 m above MSL), which is approximately 0.08 m below the current elevation of *Experiments 1 and 2*, to simulate an increase in inundation due to sea level rise (25 years at the current 3.4 mm yr⁻¹) where surface accretion would be a significant contribution to vertical change. All other

parameters reflect current conditions (suspended sediment = 25 mg L⁻¹, 44 cm tidal amplitude) or original model parameters used by Mudd et al. 2010^{35} . Finally, we calculated frontal area (λ) as a proxy for the potential of vegetation to disperse storm energy:

$$\lambda = \frac{whn}{A}$$

where w and h are the plot-mean stem width and height, n is the number of stems per plot, and A is the plot area.

Data Availability

Morphometric and derived biomass data from the experiments are posted on the Global Change Research Wetland website (http://serc.si.edu/gcrew/data) and all data are available from the corresponding authors upon request.